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## "One-Generation Reproduction Toxicity Study"

### 1. INTRODUCTORY INFORMATION

- Prerequisites

- Solid, liquid, gaseous or vapour test substance
- Chemical identification of test substance
- Purity (impurities) of test substance
- Solubility characteristics
- Melting point/boiling point (where appropriate)
- pH (where appropriate)

- Standard documents

There are no relevant international standards.

### 2. METHOD

#### A. INTRODUCTION, PURPOSE, SCOPE, RELEVANCE, APPLICATION AND LIMITS OF TEST

This Test Guideline for reproduction testing is designed to provide general information concerning the effects of a test substance on male and female reproductive performance, such as gonadal function, oestrous cycle, mating behaviour, conception, parturition, lactation and weaning. It is not designed to determine specific cause and effects in all cases and will require modifications to study substances administered by the inhalation route. The study may also provide preliminary information about developmental toxic effects of the test substance, such as neonatal morbidity, mortality, behaviour and teratogenesis and to serve as a guide for subsequent tests.

- Principle of the test method

The test substance is administered in graduated doses to several groups of males and females. Males should be dosed during growth and for at least one complete spermatogenic cycle (approximately 56 days in the mouse and 70 days in the rat) in order to elicit any adverse effects on spermeogenesis by the test substance.

Females of the P generation should be dosed for at least two complete oestrous cycles in order to elicit any adverse effects on oestrus by the test substance. The animals are then mated. The test substance is administered to both sexes during the mating period and thereafter only to females during pregnancy and for the duration of the nursing period.

## B. DESCRIPTION OF THE TEST PROCEDURE

- Preparations

Healthy young adult animals are randomised and assigned to the treatment groups. The animals are kept in cages for at least five days to allow for acclimatisation. It is recommended that the test substance be administered in the diet or drinking water. Other routes of administration are also acceptable. All animals should be dosed by the same method during the appropriate experimental period. If a vehicle or other additives are used to facilitate dosing, they should be known not to produce toxic effects. Dosing should be on a seven-day per week basis.

- Experimental animals

### *Selection of species*

This Test Guideline is designed for use with the rat or mouse. If other species are used, appropriate modifications will be necessary. Strains with low fecundity should not be used. Healthy animals, not subjected to previous experimental procedures, should be used. The test animals should be characterised as to species, strain, sex, weight and/or age.

### *Number and sex*

Each test and control group should contain a sufficient number of animals to yield about 20 pregnant females at or near term. For substances that cause sterility this may not be possible. The objective is to produce enough pregnancies and offspring to assure a meaningful evaluation of the potential of the substance to affect fertility, pregnancy and maternal behaviour in P generation animals and suckling, growth and development of the F<sub>1</sub> offspring from conception to weaning.

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### *Housing and feeding conditions*

The temperature in the experimental animal room should be 22°C ( $\pm$  3°) and the relative humidity 30 to 70 per cent. When the lighting is artificial the sequence should be 12 hours light, 12 hours dark. For feeding, conventional laboratory diets may be used with an unlimited supply of drinking water. Pregnant females should be caged individually and may be provided with nesting materials.

- Test conditions

#### *Dose levels*

At least three treatment groups and a control group should be used. If a vehicle is used in administering the test substance, the control group should receive the vehicle in the highest volume used. If a test substance causes reduced dietary intake or utilisation, then the use of a paired-fed control group may be considered necessary. Ideally, unless limited by the physical-chemical nature or biological effects of the test substance, the highest dose level should induce toxicity but not mortality in the parental P<sub>1</sub> animals.

Ideally, the intermediate dose should induce minimal toxic effects attributable to the test substance, and the low dose should not induce any observable adverse effects on the parents or offspring.

When administered by gavage or capsule the dose given each animal should be based on the individual animal's body weight and adjusted weekly for changes in body weight. For females during pregnancy, dosage may be based on daily body weight or on body weight at day 0 or 5 of pregnancy, if desired.

#### *Limit test*

In the case of substances of low toxicity as demonstrated in repeated-dose studies, if a dose of at least 1000 mg/kg produces no evidence of interference with reproductive performance, studies at other dose levels may not be considered necessary. If a preliminary study at the high dose level, with definite evidence of maternal toxicity, shows no adverse effects on fertility, studies at other dose levels may not be considered necessary.

- Performance of the test

*Experimental schedules*

Daily dosing of the parental (P) males should begin when they are about five to nine weeks old, after they have been weaned and acclimatised for at least five days. In rats dosing is continued for ten weeks prior to the mating period (for mice, eight weeks). Males should be killed and examined either at the end of the mating period or, alternatively, males may be retained on test diet for the possible production of a second litter and should be killed and examined at some time before the end of the experiment.

For parental (P) females, dosing should begin after at least five days of acclimatisation and continue for at least two weeks prior to mating. Daily dosing of the P females should continue throughout the 3-week mating period, pregnancy and up to the weaning of the F<sub>1</sub> offspring. Consideration should be given to modifications in the dosing schedule based on available information on the test substance, such as induction of its metabolism or bioaccumulation.

*Mating procedure*

Either 1:1 (one male to one female) or 1:2 (one male to two females) matings may be used in this study.

Based on 1:1 mating, one female should be placed with the same male until pregnancy occurs or three weeks have elapsed. Each morning the females should be examined for presence of sperm or vaginal plugs. Day 0 of pregnancy is defined as the day a vaginal plug or sperm are found.

Those pairs that fail to mate should be evaluated to determine the cause of the apparent infertility. This may involve such procedures as additional opportunities to mate with other proven sires or dams, microscopic examination of the reproductive organs, and examination of the oestrous cycle or spermeogenesis.

*Litter size*

Animals dosed during the fertility study are allowed to litter normally and rear their progeny to the stage of weaning without standardisation. If standardisation is carried out, the following procedure is suggested: on day 4 after birth, the size of each litter may be adjusted

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by eliminating extra pups by selection to yield, as nearly as possible, four males and four females per litter. Elimination of runts only is not appropriate. Whenever the number of male or female pups prevents having four of each sex per litter, partial adjustment (for example, five males and three females) is acceptable. Adjustments are not applicable for litters of less than eight pups.

- Observations

Throughout the test period, each animal should be observed at least once daily. Pertinent behavioural changes, signs of difficult or prolonged parturition and all signs of toxicity, including mortality, should be recorded. During pre-mating and mating periods, food consumption should be measured weekly. Optionally, during pregnancy food consumption may be measured daily. After parturition, and during lactation, food consumption measurements should be made on the same day as the litters are weighed. P males and females should be weighed on the first day of dosing and weekly thereafter. These observations should be reported individually for each adult animal.

The duration of gestation should be calculated from day 0 of pregnancy. Each litter should be examined as soon as possible after delivery to establish the number and sex of pups, stillbirths, live births and the presence of gross anomalies. Dead pups and pups killed at day 4 should be preserved and studied for possible defects.

Live pups should be counted and litters weighed on the morning after birth and on days 4 and 7 and weekly thereafter until termination of the study, when animals should be weighed individually. Physical or behavioural abnormalities observed in the dams or offspring should be recorded.

- Pathology

*Gross necropsy*

At the time of sacrifice or death during the study the animals of the P generation should be examined macroscopically for any structural abnormalities or pathological changes, with special attention paid to the organs of the reproductive system. Dead or moribund pups should be examined for defects.

***Histopathology***

The ovaries, uterus, cervix, vagina, testes, epididymides, seminal vesicles, prostate, coagulating gland, pituitary gland and target organ(s) of all P animals should be preserved for microscopic examination, if necessary. In the event that these organs have not been examined in other multiple-dose studies, they should be microscopically examined in all high-dose and control animals and in animals which die during the study, where practicable. Organs showing abnormalities in these animals should then be examined in all other P animals. In these instances microscopic examination should be made of all tissues showing gross pathological changes. As suggested under Mating procedure, above, reproductive organs of animals suspected of infertility may be subjected to microscopic examination.

**3. DATA AND REPORTING****• Treatment of results**

Data may be summarised in tabular form, showing for each test group the number of animals at the start of the test, the number of fertile males, the number of pregnant females, the types of changes and the percentage of animals displaying each type of change.

When possible, numerical results should be evaluated by an appropriate statistical method. A generally accepted statistical method should be used; the statistical methods should be selected as a part the design of the study.

**• Evaluation of results**

The findings of a reproduction toxicity study should be evaluated in terms of the observed effects, necropsy and microscopic findings. The evaluation will include the relationship between the dose of the test substance and the presence or absence, the incidence and severity, of abnormalities, including fertility, clinical abnormalities, body weight changes, effects on mortality and any other toxic effects. A properly conducted reproduction test should provide a satisfactory estimation of a no-effect level and an understanding of adverse effects on reproduction, parturition, lactation and postnatal growth.

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- Test report

The test report should also include the following information:

- species/strain used;
- toxic response data by sex and dose, including fertility, gestation, and viability indices;
- time of death during the study or whether animals survived to termination;
- table presenting the weights of each litter, the mean pup weights and the individual weights of the pups at termination;
- toxic or other effects on reproduction, offspring, postnatal growth, etc.;
- the day of observation of each abnormal litter and its subsequent course;
- body weight data for P animals;
- necropsy findings;
- a detailed description of macroscopic findings, when performed; and
- statistical treatment of results, where appropriate.

- Interpretation of results

A reproduction toxicity study will provide information on the effects of repeated oral exposure to a substance. The results of the study should be interpreted in conjunction with the findings of subchronic, teratogenic and other studies. Extrapolation of the results of the study to man is valid to a limited degree, although it can provide useful information on no-effect levels and permissible human exposure.

#### **4. LITERATURE**

1. E.I. Goldenthal, *Guidelines for Reproduction Studies for Safety Evaluation of Drugs for Human Use*. Drug Review Branch, Division of Toxicological Evaluation, Bureau of Science, Federal Drug Administration, Washington, D.C. (1966).
2. T. Hasegawa, M. Hayashi, F.J.G. Ebling and I.W. Henderson, *Fertility and Sterility*. Elsevier, New York (1973).

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3. E.F. Oakberg, *Am. J. Anat.* 99, 507-516 (1956).
4. E.C. Roosen-Runge, *Biol. Rev.* 37, 343-377 (1962).
5. R.D. Yates and M. Gordon, *Male Reproductive System*. Masson, New York (1977).
6. National Academy of Sciences, *Principles and Procedures for Evaluating the Toxicity of Household Substances* (Washington, 1977).
7. A.K. Palmer, in *Handbook of Teratology*, Vol. 4 (edited by J.G. Wilson and J. Clarke Graser) Plenum Press, New York (1978).
8. A.K. Palmer, in *Developmental Toxicology* (edited by C.A. Kimmel and B. Buelke-Sam) Raven Press, New York (1981).

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